

## Model repositories

Name	Acronym	Website	Country
Rat Resource Center	RRC	<a href="http://www.rrrc.us/search/?strainStatus=available">http://www.rrrc.us/search/?strainStatus=available</a>	USA
National BioResource Project	NBRP	<a href="http://www.anim.med.kyoto-u.ac.jp/NBR/strainsx/Strains_list.aspx">http://www.anim.med.kyoto-u.ac.jp/NBR/strainsx/Strains_list.aspx</a>	Japan

## Reporter rat models

LE, Long Evans; SD, Sprague-Dawley; DA, Dark Agouti; LEW, Lewis; F344, Fischer  
L, alive animals; CP, cryopreserved (sperm and/or embryo)

System/Tissue	Promotor / Reporter gene	Model name: strain-mutation type(promoter-reporter gene)	Phenotype Specificity	Reference / Donor (Lab)	Depository ID#	L/CP
Neuronal	cFos (sp)/GFP	LE-Tg (cFos-eGFP)2Ottc	Expression in cells expressing immediate early gene Fos (prefrontal cortex, dorsal striatum, hippocampus)	B.Harvey-J. Pickel (NIDA/NIMH)	RRRC #00766	CP
	S-100b (sp)/GFP	Wistar- Tg(S-100b-GFP)	High expression-Specifically in pituitary folliculo-stellate cells, brain astrocytes, adipocytes, chondrocytes	[1-3]	NBRP Rat N#0371	L/CP
	Gnrh1 (sp)/GFP	Wistar-Tg(Gnrh1-EGFP)	EGFP fluorescence is observed only in Gnrh1-immunoreactive neurons, approximately one third of which has strong EGFP fluorescence.	[4-11]	NBRP Rat #0469	L/CP
	Nestin enhancer / Hspa1b promoter / GFP	SD-Tg(Nes-Hspa1b-EGFP)		[12]	NBRP Rat #0352	CP
	Tub $\beta$ 1a / GFP	SD-Tg(Tuba1a-EYFP)		[13]	NBRP Rat #0353	CP
Salivary gland Lung	Aqp5 (sp) / GFP	SD-Tg(Aqp5-GFP)	Expression in salivary gland (acini cells) in alveolar epithelial type I (AT1) cells in the lung.	[14]	RRRC #00427	CP
	Plcb2 (sp)/GFP	Wistar-Tg(Plcb2-WGA,-EGFP)	GFP are expressed in almost all PLCb2-positive cells in the taste buds.	Keiko Abe	NBRP Rat #0214	CP
Germ cells	Pou5f1 (sp) / GFP	SD-Tg(Pou5f1-EGFP)	Expression in germ cells	C. Dann (U. Texas Southwestern Medical Center at Dallas)	RRRC #00319	CP
	Pou5f1 (sp)/ DsRed	SD-Tg(Pou5f1-Dsred)	DsRed expression in inner cell mass of blastocyst stage.	C. Dann	RRRC #00320	CP

<b>Endocrine</b>	RIP7 (sp)/GFP	SD-Tg(RIP7-RLuc-YFP)	Expression in mature pancreatic endocrine beta (B)-cells.	Vincent Poitout (CRCHUM)	RRTC #00757	CP
<b>Liver</b>	Alb (sp)/ DsRed	Wistar-Tg(Alb-DsRed2)	Ds red expression is seen in the liver of fetuses for both sexes. Ds red expression is seen in the liver of young adult males but not in the liver of young adult females.	[15]		CP
<b>Ubiquitous</b>	mCMV/TRE /GFP	SD-Tg (mCMV/TRE-eGFP)	Dox-controlled ubiquitous expression	Y.Agca (U.Missouri)		CP
	CAG /GFP	LEW-Tg(CAG-EGFP)	Ubiquitous eGFP expression	E. Kobayashi (Jichi Medical School)		L/CP
	CAG / GFP	LEW-Tg (CAG-GFP)	High Ubiquitous expression and in cultured-neurospheres	[16,17]		
	CAG /GFP	Wistar-TgN(CAG-GFP)	Ubiquitous eGFP expression	[18,19]		CP
	CAG /GFP	Wistar-Tg(CAG-GFP)	Moderate to high ubiquitous expression	[18]		
	CAG /GFP	Wistar-Tg(CAG-GFP)	Weak to strong Dependent of developmental stage	[20]		
	CAG /GFP	Wistar-Tg(CAG-GFP)	Moderate expression in Kidney, Testis, Lung, Spleen, some cerebral area, spinal cord and cultured-neurospheres	[21]		
	CAG / GFP	SD-Tg(CAG-EGFP)		Masaru Okabe Osaka University		L
		SD-KI Rosa26-GFP		[22]		
		DA/Wistar-KI Rosa26-tdTomato		[23]		
		DA/KI Sox10-dsRed	Neonatal brain ; oligodendrocyte lineage	[24]		
		Wistar/Rosa26-KI H2B-tdTomato	oocyte nuclei (MII phase; PN; from 2c up to blastocyste)	[25]		
	UbC / GFP	SD-Tg(UbC-eGFP/ Dazl-shRNA)	knockdown of the Dazl gene. GFP expression in males but silenced in females (unknown reasons)	[26]		CP
	UbC / GFP	F344-Tg(UbC-eGFP)	Ubiquitous eGFP expression	[27]		L/CP
	UbC / GFP	SD-Tg (UbC /WRE-GFP)	Weak and variable between individual rats Kidney, Testis, Lung, Spleen, some cerebral area, spinal cord	[21]		
	Pgk / GFP	SD-Tg (PGK-GFP)	High to low expression	[28]		L/CP
Rosa / Luciferase	LEW-Tg(Rosa-luc)		[29]			
HIV / LacZ	SD-Tg(HIV-LacZ)	HIV-LacZ transgenic rats display immune tolerance towards B-galactosidase so that LacZ expression is not silenced. Ubiquitous LacZ tissue expressio	[30,31]		CP	

- [1] E. Itakura, K. Odaira, K. Yokoyama, M. Osuna, T. Hara, K. Inoue, Generation of transgenic rats expressing green fluorescent protein in S-100beta-producing pituitary folliculo-stellate cells and brain astrocytes, *Endocrinology*. 148 (2007) 1518–1523. <https://doi.org/10.1210/en.2006-1390>.
- [2] R. Syaidah, T. Tsukada, M. Azuma, K. Horiguchi, K. Fujiwara, M. Kikuchi, T. Yashiro, Fibromodulin Expression in Folliculostellate Cells and Pericytes Is Promoted by TGF $\beta$  Signaling in Rat Anterior Pituitary Gland, *Acta Histochem. Cytochem.* 49 (2016) 171–179. <https://doi.org/10.1267/ahc.16021>.
- [3] K.C. Patterson, V.E. Hawkins, K.M. Arps, D.K. Mulkey, M.L. Olsen, MeCP2 deficiency results in robust Rett-like behavioural and motor deficits in male and female rats, *Hum. Mol. Genet.* 25 (2016) 3303–3320. <https://doi.org/10.1093/hmg/ddw179>.
- [4] M. Kato, K. Ui-Tei, M. Watanabe, Y. Sakuma, Characterization of voltage-gated calcium currents in gonadotropin-releasing hormone neurons tagged with green fluorescent protein in rats, *Endocrinology*. 144 (2003) 5118–5125. <https://doi.org/10.1210/en.2003-0213>.
- [5] C. Yin, H. Ishii, N. Tanaka, Y. Sakuma, M. Kato, Activation of A-type gamma-amino butyric acid receptors excites gonadotrophin-releasing hormone neurones isolated from adult rats, *J. Neuroendocrinol.* 20 (2008) 566–575. <https://doi.org/10.1111/j.1365-2826.2008.01697.x>.
- [6] M. Kato, N. Tanaka, S. Usui, Y. Sakuma, The SK channel blocker apamin inhibits slow afterhyperpolarization currents in rat gonadotropin-releasing hormone neurones, *J. Physiol.* 574 (2006) 431–442. <https://doi.org/10.1113/jphysiol.2006.110155>.
- [7] M. Watanabe, Y. Sakuma, M. Kato, GABAA receptors mediate excitation in adult rat GnRH neurons, *Biol. Reprod.* 81 (2009) 327–332. <https://doi.org/10.1095/biolreprod.108.074583>.
- [8] M. Kato, N. Tanaka, H. Ishii, C. Yin, Y. Sakuma, Ca $^{2+}$  channels and Ca $^{2+}$ -activated K $^{+}$  channels in adult rat gonadotrophin-releasing hormone neurones, *J. Neuroendocrinol.* 21 (2009) 312–315. <https://doi.org/10.1111/j.1365-2826.2009.01849.x>.
- [9] N. Tanaka, H. Ishii, C. Yin, M. Koyama, Y. Sakuma, M. Kato, Voltage-gated Ca $^{2+}$  channel mRNAs and T-type Ca $^{2+}$  currents in rat gonadotropin-releasing hormone neurons, *J. Physiol. Sci. JPS.* 60 (2010) 195–204. <https://doi.org/10.1007/s12576-010-0085-z>.
- [10] M. Koyama, C. Yin, H. Ishii, Y. Sakuma, M. Kato, Somatostatin inhibition of GnRH neuronal activity and the morphological relationship between GnRH and somatostatin neurons in rats, *Endocrinology*. 153 (2012) 806–814. <https://doi.org/10.1210/en.2011-1374>.
- [11] H. Tada, Y. Kuroki, T. Funabashi, Y. Kamiya, T. Goto, K. Suyama, A. Sano, D. Mitsushima, A.M. Etgen, T. Takahashi, Phasic synaptic incorporation of GluR2-lacking AMPA receptors at gonadotropin-releasing hormone neurons is involved in the generation of the luteinizing hormone surge in female rats, *Neuroscience*. 248 (2013) 664–669. <https://doi.org/10.1016/j.neuroscience.2013.06.040>.
- [12] K. Sawamoto, N. Nakao, K. Kakishita, Y. Ogawa, Y. Toyama, A. Yamamoto, M. Yamaguchi, K. Mori, S.A. Goldman, T. Itakura, H. Okano, Generation of dopaminergic neurons in the adult brain from mesencephalic precursor cells labeled with a nestin-GFP transgene, *J. Neurosci. Off. J. Soc. Neurosci.* 21 (2001) 3895–3903.
- [13] Y. Ogawa, K. Sawamoto, T. Miyata, S. Miyao, M. Watanabe, M. Nakamura, B.S. Bregman, M. Koike, Y. Uchiyama, Y. Toyama, H. Okano, Transplantation of in vitro-expanded fetal neural progenitor cells results in neurogenesis and functional recovery after spinal cord contusion injury in adult rats, *J. Neurosci. Res.* 69 (2002) 925–933. <https://doi.org/10.1002/jnr.10341>.
- [14] B. Zhou, D.K. Ann, P. Flodby, P. Minoo, J.M. Liebler, E.D. Crandall, Z. Borok, Rat aquaporin-5 4.3-kb 5'-flanking region differentially regulates expression in salivary gland and lung in vivo, *Am. J. Physiol. Cell Physiol.* 295 (2008) C111-120. <https://doi.org/10.1152/ajpcell.90620.2007>.
- [15] Y. Sato, Y. Igarashi, Y. Hakamata, T. Murakami, T. Kaneko, M. Takahashi, N. Seo, E. Kobayashi, Establishment of Alb-DsRed2 transgenic rat for liver regeneration research, *Biochem. Biophys. Res. Commun.* 311 (2003) 478–481. <https://doi.org/10.1016/j.bbrc.2003.09.230>.
- [16] H. Inoue, I. Ohsawa, T. Murakami, A. Kimura, Y. Hakamata, Y. Sato, T. Kaneko, M. Takahashi, T. Okada, K. Ozawa, J. Francis, P. Leone, E. Kobayashi, Development of new inbred transgenic strains of rats with LacZ or GFP, *Biochem. Biophys. Res. Commun.* 329 (2005) 288–295. <https://doi.org/10.1016/j.bbrc.2005.01.132>.
- [17] J.S. Francis, A. Olariu, E. Kobayashi, P. Leone, GFP-transgenic Lewis rats as a cell source for oligodendrocyte replacement, *Exp. Neurol.* 205 (2007) 177–189. <https://doi.org/10.1016/j.expneurol.2007.01.039>.
- [18] Y. Hakamata, K. Tahara, H. Uchida, Y. Sakuma, M. Nakamura, A. Kume, T. Murakami, M. Takahashi, R. Takahashi, M. Hirabayashi, M. Ueda, I. Miyoshi, N. Kasai, E. Kobayashi, Green fluorescent protein-transgenic rat: a tool for organ transplantation research, *Biochem. Biophys. Res. Commun.* 286 (2001) 779–785. <https://doi.org/10.1006/bbrc.2001.5452>.
- [19] M. Hirabayash, M. Kato, T. Aoto, A. Sekimoto, M. Ueda, I. Miyoshi, N. Kasai, S. Hochi, Offspring derived from intracytoplasmic injection of transgenic rat sperm, *Transgenic Res.* 11 (2002) 221–228. <https://doi.org/10.1023/a:1015210604906>.

- [20] K. Takeuchi, A. Sereemasapun, T. Inagaki, Y. Hakamata, T. Kaneko, T. Murakami, M. Takahashi, E. Kobayashi, S. Ookawara, Morphologic characterization of green fluorescent protein in embryonic, neonatal, and adult transgenic rats, *Anat. Rec. A. Discov. Mol. Cell. Evol. Biol.* 274 (2003) 883–886. <https://doi.org/10.1002/ar.a.10111>.
- [21] A.J. Mothe, I. Kulbatski, R.L. van Bendegem, L. Lee, E. Kobayashi, A. Keating, C.H. Tator, Analysis of green fluorescent protein expression in transgenic rats for tracking transplanted neural stem/progenitor cells, *J. Histochem. Cytochem. Off. J. Histochem. Soc.* 53 (2005) 1215–1226. <https://doi.org/10.1369/jhc.5A6639.2005>.
- [22] S. Remy, L. Tesson, S. Menoret, C. Usal, A. De Cian, V. Thepenier, R. Thinard, D. Baron, M. Charpentier, J.-B. Renaud, R. Buelow, G.J. Cost, C. Giovannangeli, A. Fraichard, J.-P. Concordet, I. Anegon, Efficient gene targeting by homology-directed repair in rat zygotes using TALE nucleases, *Genome Res.* 24 (2014) 1371–1383. <https://doi.org/10.1101/gr.171538.113>.
- [23] T. Kobayashi, M. Kato-Itoh, T. Yamaguchi, C. Tamura, M. Sanbo, M. Hirabayashi, H. Nakauchi, Identification of Rat Rosa26 Locus Enables Generation of Knock-In Rat Lines Ubiquitously Expressing tdTomato, *Stem Cells Dev.* 21 (2012) 2981–2986. <https://doi.org/10.1089/scd.2012.0065>.
- [24] Y. Chen, S. Spitzer, S. Agathou, R.T. Karadottir, A. Smith, Gene Editing in Rat Embryonic Stem Cells to Produce In Vitro Models and In Vivo Reporters, *Stem Cell Rep.* 9 (2017) 1262–1274. <https://doi.org/10.1016/j.stemcr.2017.09.005>.
- [25] T. Goto, T. Kobayashi, H. Hara, M. Sanbo, S. Hochi, H. Nakauchi, M. Hirabayashi, Knock-in of a histone H2B-tdTomato reporter into the Rosa26 locus allows visualization of cell nuclei in rats, *Mol. Reprod. Dev.* 82 (2015) 916–917. <https://doi.org/10.1002/mrd.22584>.
- [26] C.T. Dann, A.L. Alvarado, R.E. Hammer, D.L. Garbers, Heritable and stable gene knockdown in rats, *Proc. Natl. Acad. Sci. U. S. A.* 103 (2006) 11246–11251. <https://doi.org/10.1073/pnas.0604657103>.
- [27] C. Lois, E.J. Hong, S. Pease, E.J. Brown, D. Baltimore, Germline transmission and tissue-specific expression of transgenes delivered by lentiviral vectors, *Science.* 295 (2002) 868–872. <https://doi.org/10.1126/science.1067081>.
- [28] S. Remy, L. Tesson, C. Usal, S. Menoret, V. Bonnamain, V. Nerriere-Daguin, J. Rossignol, C. Boyer, T.H. Nguyen, P. Naveilhan, L. Lescaudron, I. Anegon, New lines of GFP transgenic rats relevant for regenerative medicine and gene therapy, *Transgenic Res.* 19 (2010) 745–763. <https://doi.org/10.1007/s11248-009-9352-2>.
- [29] Y. Hakamata, T. Murakami, E. Kobayashi, “Firefly rats” as an organ/cellular source for long-term in vivo bioluminescent imaging, *Transplantation.* 81 (2006) 1179–1184. <https://doi.org/10.1097/01.tp.0000203137.06587.4a>.
- [30] S. Ménoret, D. Aubert, L. Tesson, C. Braudeau, V. Pichard, N. Ferry, I. Anegon, lacZ transgenic rats tolerant for beta-galactosidase: recipients for gene transfer studies using lacZ as a reporter gene, *Hum. Gene Ther.* 13 (2002) 1383–1390. <https://doi.org/10.1089/104303402760128603>.
- [31] D. Aubert, S. Ménoret, E. Chiari, V. Pichard, S. Durand, L. Tesson, P. Moullier, I. Anegon, N. Ferry, Cytotoxic immune response blunts long-term transgene expression after efficient retroviral-mediated hepatic gene transfer in rat, *Mol. Ther. J. Am. Soc. Gene Ther.* 5 (2002) 388–396. <https://doi.org/10.1006/mthe.2002.0561>.